

Protein Collision on DNA

Ja Yil Lee, Assist. Prof. School of Life Sciences, UNIST

July 09, 2018



2018 Summer School for Biophysics in Postech

Outline

- I. Introduction and background
- II. DNA curtain assay
- **III.** Protein movement and collision on DNA
- **IV.** Summary
- V. Beyond DNA curtain

Biophysics

- Biophysics is not easy because it contains a wide range of academic areas
- Physicists encounter huge barriers to study biology, and biologists feel frustrated when they study physics!
 - \rightarrow Hard to get familiar with the other discipline
- Studying biophysics is like a spiral staircase
 - While you are walking on the step, it seems that you are just rotating around the same path. But at a certain time, you can get to higher position and see a different view



Biophysics

What is the biophysics?

Biophysics Society says

"Biophysics is a bridge between biology and physics"

"Biology studies life in its variety and complexity. It describes how organisms go about getting food, communicating, sensing the environment, and reproducing. On the other hand, physics looks for mathematical laws of nature and makes detailed predictions about the forces that drive idealized systems. *Spanning the distance between the complexity of life and the simplicity of physical laws is the challenge of biophysics*. Looking for the patterns in life and analyzing them with math and physics is a powerful way to gain insights."

Wiki says

"Biophysics or biological physics is an interdisciplinary science that *applies the approaches and methods of physics to study biological systems*. Biophysics covers all scales of biological organization, from molecular to organismic and populations. Biophysical research shares significant overlap with biochemistry, physical chemistry, nanotechnology, bioengineering, computational biology, biomechanics, and systems biology."

Biophysics



Collision



Biological Background

DNA

Cell



Base-pair (bp) A-T and G-C

> Double-stranded Helical structure (B-DNA)

Significance of Protein Collision



Limit of Traditional Biological Methods

- Traditional biological techniques
 - 1. Use a large amount of biomolecules in test tubes
 - 2. Measure the average effect of biomolecules or biomolecular reactions

- Limitation of traditional methods
 - 1. Cannot reveal the details of biomolecular properties and interactions
 - 2. Cannot observe the biomolecular movement
 - 3. Not easy to obtain precise kinetic information of biomolecular reactions

✓ New technology is demanded to overcome the limitations
 → Single-molecule biophysics is the solution!!!

Single-Molecule Biophysics

Observe the behavior of individual bio-molecules with the help of advanced physical techniques

> Advantages

- 1. Exclude the *ensemble average effect* of massive samples
 - \rightarrow Reveal the hidden sub-states of biomolecules
- 2. Measure the kinetics of molecular reactions more precisely
 - \rightarrow No need for the stop flow
- 3. Mechanically manipulate biomolecules and examine the response to the mechanical stress
 - \rightarrow Apply force and torque to biomolecules
- 4. Observe biomolecular interaction in more direct way
 - \rightarrow Directly visualize the molecular motion and interactions

Disadvantages

- 1. Gather sufficient number of data for statistical reliability
- 2. Modify biomolecules

Single-Molecule Biophysics

Ensemble view

Single-molecule view





Beginning of Single-Molecule Spectroscopy

VOLUME 62, NUMBER 21

PHYSICAL REVIEW LETTERS

Optical Detection and Spectroscopy of Single Molecules in a Solid

W. E. Moerner and L. Kador^(a)

IBM Research Division, Almaden Research Center, San Jose, California 95120 (Received 17 March 1989)

Using two different double-modulation techniques, we have observed the optical-absorption spectrum of single dopant molecules of pentacene in a *p*-terphenyl host crystal at liquid-helium temperatures. To achieve this, frequency-modulation spectroscopy was combined either with Stark or ultrasonic modulation to remove interfering background signals from residual amplitude modulation, and the number of molecules in resonance was reduced to one by operating in the wings of the inhomogeneous line. Triplet bottleneck saturation appears to be suppressed in the single-molecule regime.

PACS numbers: 78.50.-w, 33.20.Kf, 33.70.Jg

W. E. Moerner won the Nobel prize in chemistry 2014!

Single-Molecule Techniques: Fluorescence

Single-molecule FRET

FCS (Florescence correlation spectroscopy)



Time



Single particle tracking



Single-Molecule Techniques: Force



Single-Molecule Techniques: Electronic Signal

Carbon nanotube device

ELECTRONIC SIGNAL

Membrane protein

Open Close Time



2014 Nobel Prize in Chemistry Super-Resolution Microscopy



Eric Betzig

Stephan W. Hell

William E. Moerner

DNA Curtain Assay

DNA Curtain





Double-Tethered DNA Curtain









Criss-cross

Separate curtain



AFM image





DNA

intersections

Single-Stranded DNA Curtain





Single-Stranded DNA Curtains



Anal. Chem. 84, 7607 (2012)

Single-Stranded DNA Curtains





Protein Movement and Collision

DNA Motor Protein: FtsK









KOPS-Guided Translocation of FtsK



CHS Perspectives

1. Single FtsK Motion on DNA



FtsK Binding without ATP

Green: DNA, magenta: FtsK





No ATP

Mutant FtsK & ATP

PNAS 109, 6531 (2012)

FtsK Translocation



KOPS-Guided Movement

- FtsK does not recognize KOPS during translocation.
 Hypothesis: initial binding on KOPS determines the orientation of FtsK movement.
- **ATP-chase:** place FtsK on KOPS with ADP, then inject ATP into the flowcell.



Lee et al. PNAS (2012)

KOPS guides the orientation of FtsK only when the FtsK departs from KOPS !!!

KOPS-Guided Initiation



2. Collisions with Stationary Protein Obstacles



Collisions with DNA Binding Proteins

EcoRI^{E111Q}



PDB: 2ckq

Mutant FtsK



http://www.bioch.ox.ac.uk



PDB: 1LBG



RNAP

Curr Opin Struct Biol. 11, 155 (2001)



Tus

Cell 125, 1309 (2006)

FtsK Collision with EcoRIE111Q

Green: FtsK, Magenta: EcoRIE111Q



Collisions with Other Protein Roadblocks

Green: FtsK, Magenta: roadblocks



Collision Outcome for All Protein Roadblocks



Lee et al. Mol Cell (2014)

Mechanism of Bypass (or Reverse)



FtsK hexamer is wrapping dsDNA.

 \rightarrow How can the FtsK bypass the protein roadblock?

Dissociation & re-association mechanism



*k*_{off} (*K*_D) vs. Push Probability



Mol Cell 54, 832 (2014)

Collision with Multiple FtsK Hexamers

- *In vivo,* FtsK hexamers are localized at the septum.
 - \rightarrow The local concentration of FtsK hexamers must be fairly high.
 - \rightarrow Multiple FtsK hexamers seem to work on the chromosomal DNA simultaneously.



Multiple FtsK hexamers can remove protein roadblocks!!!

Translocation Mechanism through Crowded DNA



3. Collision with Another Moving Protein



Collision with Another DNA Translocase (RecBCD)



Collision of FtsK and RecBCD

Green: FtsK, magenta: RecBCD



Summary

Using the single-molecule DNA curtains technique, we directly visualize the behavior of FtsK and its collision with other proteins.

- FtsK is assembled into a hexamer on KOPS, which guides FtsK orientation at the departure.
- FtsK translocates on DNA rapidly and processively.
- FtsK can continue to translocate by bypassing or evicting protein obstacles.
- Chemical constants (K_D or k_{off}) are highly related to the mechanical binding strength of DNA-binding proteins.
- Specific interaction between γ -subdomain and XerD prevents displacement of XerCD from the *dif* site.
- RecBCD is more powerful than FtsK although FtsK is faster than RecBCD.

Beyond DNA Curtain

DNA Curtains + Optical Tweezers



BBRC 426, 565 (2012)



Multiple Optical-Trap & DNA Curtains





Actin & Actin Filaments

- Globular protein (42 kDa).
- Highly conserved and abundant in eukaryotic cells.
- Monomeric form (G-actin) and filamentous form (F-actin).
- Polarized polymerization: barbed (+) end is faster than pointed (-) end.



Biological Functions of Actin Filament

Cytoskeletal network



Cytokinesis: contractile ring





Nature Reviews | Molecular Cell Biology





Cellular transport



Actin Curtain



Polymerization of single actin filament



Acknowledgement

Prof. Eric C. Greene (Columbia Univ)

Zhi Qi Tsuyoshi Terakawa Sy Redding Bryan Gibb Justin Steinfeld

Johannes Stigler Fabian Erdel Luisina de Tullio Kyle Kaniecki Frank Ma Daniel Duzdevich Corentin Moevus Myles Marshell Patrick Sung's Lab (Yale Univ)

> Youngho Kwon Hengyao Niu Xiaoyu Xue Will Gaines

Prof. Thomas Pollard (Yale Univ)



Prof. Ilya J. Finkelstein (Univ Texas)

Prof. David Sherratt (Oxford Univ)

Lidia Arcoszewsla Estelle Crozat HHHMI HOWARD HUGHES MEDICAL INSTITUTE

Our laboratory is always open!

Please knock on the door of my laboratory!

Questions?